

Microencapsulation Technology

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INTRODUCTION

Microencapsulation technology has been used from 1930s in packaging flavors and vitamins. Since the first commercial product was introduced for the carbonless copying paper,^[1] the technology has advanced to a new level. Various microencapsulation techniques are available nowadays, and the microencapsulated products are widely used in pharmaceutical, biomedical, agricultural, food, consumer products, and cosmetic industries. Representative applications of microparticles in the pharmaceutical and biomedical industries include:

- Taste and odor masking^[2]
- Protection of drugs from the environment^[3]
- Particle size reduction for enhancing solubility of the poorly soluble drugs^[4]
- Sustained or controlled drug delivery^[5]
- Cell encapsulation^[6]

The microparticle system has become an indispensable part of the controlled drug delivery fields for the past few decades since it can readily be adapted for various administration methods. In particular, biodegradable polymeric microparticles can provide a number of advantages over conventional parenteral formulations:

- **Sustained delivery:** By encapsulating a drug in a polymer matrix, which limits access of the biological fluid into the drug until the time of degradation, microparticles maintain the blood level of the drug within a therapeutic window for a prolonged period. Toxic side effects can be minimized, and patient compliance can be improved by reducing the frequency of administration.
- **Local delivery:** Subcutaneously or intramuscularly applied microparticles can maintain a therapeutically effective concentration at the site of action for a desirable duration. The local delivery system obviates systemic drug administration for local therapeutic effects and can reduce the related

systemic side effects. This system has proven beneficial for delivery of local anesthetics.^[7]

- **Pulsatile delivery:** While burst and pulsatile release is not considered desirable for the sustained delivery application, this release pattern proves to be useful for delivery of antibiotics and vaccines. Pulsatile release of antibiotics can alleviate evolution of the bacterial resistance. In the vaccine delivery, initial burst followed by delayed release pulses can mimic an initial and boost injection, respectively.^[8]

With the recent advance of biotechnology and polymer chemistry, the use of microparticle systems will continue to grow for a variety of applications. The objective of this article is to provide a review of the technical aspects of the microencapsulation techniques that have been widely used in the pharmaceutical industry and recent advances of the technology so that the pharmaceutical scientists can take full advantage of the existing assets of this area in developing new microparticle systems.

TERMINOLOGY

The microencapsulation processes produce small particles ranging in size from 1 to 1000 μm . There are different names for these particles: microparticle, microsphere, microcapsule, and micromatrix. Although they are often used interchangeably, distinctions can be made such that microcapsules are made of one or multiple core substances (solid or liquid) that are surrounded by a distinct capsule wall, whereas micromatrices are polymeric matrices in which the encapsulated substances are homogeneously dispersed. Microparticles or microspheres are general terminologies that involve both. Although micromatrices are also called microspheres depending on the authors,^[9] we will follow the former definition in this chapter. In consideration of the scope of this chapter, current discussion is limited to the microparticles that utilize natural or synthetic polymers as an encapsulating material.

MICROENCAPSULATION TECHNIQUES

Existing microencapsulation techniques have been reviewed extensively,^[5,9-12] and for this reason, here we will briefly summarize representative microencapsulation techniques.

Coacervation

The coacervation method is one of the earliest microencapsulation techniques, which has been used for various consumer products. This method is based on separation of a solution of hydrophilic polymer(s) into two phases, which are small droplets of a dense polymer-rich phase and a dilute liquid phase. Coacervation can be divided into simple and complex coacervation depending on the number of polymers that are involved in the formation of microparticles.

Simple coacervation

This process involves only one polymer (e.g., gelatin, polyvinyl alcohol, carboxymethyl cellulose), and the phase separation can be induced by conditions that result in desolvation (or dehydration) of the polymer phase. These conditions include addition of a water-miscible non-solvent, such as ethanol, acetone, dioxane, isopropanol, or propanol,^[13] addition of inorganic salts, such as sodium sulfate,^[14] and temperature change.^[15]

Complex coacervation

This process involves two hydrophilic polymers of opposite charges.^[13] Neutralization of the overall positive charges on one of the polymers by the negative

charge on the other is used to bring about separation of the polymer-rich phase (Fig. 1). The best-known example is the gelatin-gum arabic system pioneered by Bungenberg de Jong in the early 1940s.^[16] Since electrostatic interactions are involved, the pH of the medium is very important. For example, in the gelatin-gum arabic system, pH should be below the isoelectric point of gelatin so that the gelatin can maintain the positive charge. Once embryonic coacervates form around the dispersed oil or solid phases, these polymer complexes are stabilized by cross-linking using glutaraldehyde.

Commercial products

The first commercial microparticle product based on the complex coacervation method was carbonless copy paper developed by National Cash Register Corp.^[17] The back side of the first page is coated with microcapsules in the 3–10 μm size range made of a gelatin-gum arabic shell by the coacervation technique. In the center of the capsules is the oil containing colorless color-forming agent (e.g., crystal violet lactone). The front side of the second page is coated with a developing layer. The pressure imposed on both sheets of paper upon writing induces breakage of the microcapsules and makes the colorless color-forming agent released and react with the developing layer to develop color. The microcapsules have also been used in Scratch-N-Sniff[®] scent strips and Snap-N-Burst[®] fragrance samplers.

Emulsion Solidification

Microparticles can be produced from emulsion of two or more immiscible liquids. For example, a

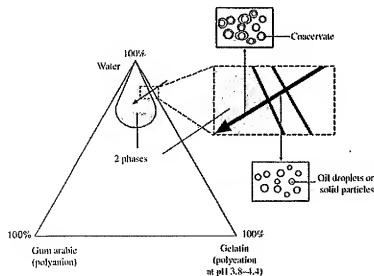


Fig. 1 Phase diagram for complex coacervation.

hurdles are maintaining the stability of encapsulated drugs throughout the lifetime of the products, manipulating release rates according to the applications, and transferring bench scale processes to the manufacturing scale. Some of the answers to those problems have been provided by advances in polymer chemistry, formulation efforts, and recent progresses in new microencapsulation techniques. The microencapsulation technology will remain as one of the most important areas in drug delivery and various other applications.

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ARTICLE OF FURTHER INTEREST

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REFERENCES

- Green, B.K.; Schleicher, L. Oil-containing Microscopic Capsules and Method of Making Them. U.S. Patent 2809457, 1957.
- Bakon, J.A.; Powell, T.C.; Szotak, P.S. Recent advances using microencapsulation for taste-masking of bitter drugs. In *Microencapsulation and Nanoparticles in Medicine and Pharmacy*; Donbrow, M., Ed.; CRC Press: Boca Raton, FL, 1992; 149-156.
- Arifa, B.; Arifa, M.Y.; Kas, H.S.; Hincal, A.A.; Hasirci, V. In-vitro studies of enteric coated diclofenac sodium carboxymethyl cellulose microspheres. *J. Microencapsul.* 1996, 13 (6), 689-699.
- Müller, R.H.; Peters, K. Nanosuspensions for the formulation of poorly soluble drugs. I. Preparation by a size-reduction technique. *Int. J. Pharm.* 1998, 160 (2), 229-237.
- Mathiowetz, E.; Kreitz, M.R. Microencapsulation. In *Encyclopedia of Controlled Drug Delivery*; Mathiowetz, E., Ed.; John Wiley & Sons, Inc.: New York, U.S.A., 1999; Vol. 2, 493-546.
- Lim, F.; Sun, A.M. Microencapsulated islets as bioartificial endocrine pancreas. *Science* 1980, 210, 908-910.
- Kohane, D.S.; Lipp, M.; Kinney, R.C.; Lotan, N.; Langer, R. Sciatic nerve blockade with lipid-protein-sugar particles containing bupivacaine. *Pharm. Res.* 2000, 17 (10), 1243-1249.
- Ying, M.; Thomasin, C.; Merkle, H.P.; Gander, B.; Corradin, G. A single administration of tetanus toxoid in biodegradable microspheres elicits T cell and antibody responses similar or superior to those obtained with aluminum hydroxide. *Vaccine* 1995, 13 (7), 683-689.
- Yeo, Y.; Back, N.; Park, K. Microencapsulation methods for delivery of protein drugs. *Biotechnol. Bioprocess Eng.* 2001, 6 (4), 213-230.
- Benoit, J.-P.; Marchais, H.; Rolland, H.; Velde, V.V. Biodegradable microspheres: advances in production technology. In *Microencapsulation: Methods and Industrial Application*; Benita, S., Ed.; Marcel Dekker, Inc.: New York, U.S.A., 1996; Vol. 73, 35-72.
- Thies, C. A survey of microencapsulation processes. In *Microencapsulation: Methods and Industrial Applications*; Benita, S., Ed.; Marcel Dekker, Inc.: New York, U.S.A., 1996; Vol. 73, 1-19.
- Gouin, S. Microencapsulation industrial appraisal of existing technologies and trends. *Trends Food Sci. Technol.* 2004, 15 (7-8), 330-347.
- Mohanty, B.; Bohidar, H.B. Systematic of alcohol-induced simple concentration in aqueous gelatin solutions. *Biocromolecules* 2003, 4 (4), 1080-1086.
- Weiss, G.; Knoch, A.; Laicher, A.; Stanislaus, P.; Daniels, R. Simple conservation of hydroxypropyl methyl cellulose phthalate (HPMCP). I. Temperature and pH dependency of coacervate formation. *Int. J. Pharm.* 1995, 124 (1), 87-96.
- Purgas, D.J.; Singh, O. Spontaneous formation of small sized albumin/lecithin coacervate particles. *J. Pharm. Pharmacol.* 1993, 45, 586-591.
- de Jong, H.G.B. Complex colloid systems (Chapter X). In *Colloid Science*; Elsevier: New York, 1949.
- Sah, H. Microencapsulation techniques using ethyl acetate as a dispersed solvent; effects of its extraction rate on the characteristics of PLGA microspheres. *J. Controlled Release* 1997, 47 (3), 233-245.
- Herrmann, J.; Bodmeier, R. Somatostatin containing biodegradable microspheres prepared by a modified solvent evaporation method based on W/O/W-multiple emulsions. *Int. J. Pharm.* 1995, 126 (1-2), 129-138.
- Bodmeier, R.; McGinity, J.W. Solvent selection in the preparation of PLA microspheres prepared by the solvent evaporation method. *Int. J. Pharm.* 1988, 43, 179-186.
- Yeo, Y.; Park, K. Control of encapsulation efficiency and initial burst in polymeric microparticle systems. *Arch. Pharmacol. Res.* 2004, 27 (1), 1-12.
- Kempen Diederik, H.R.; Lu, L.; Zhu, X.; Kim, C.; Jabbari, E.; Dhert Wouter, J.A.; Currier Bradford, L.; Yaszemski Michael, J. Development of biodegradable poly(propylene fumarate)/poly(lactic-co-glycolic acid) blend microspheres. II. Controlled drug release and microsphere degradation. *J. Biomed. Mater. Res.* 2004, 70A (2), 293-302.
- Dittrich, M.; Hampf, J.; Soukup, F. Branched oligoester microspheres fabricated by a rapid emulsion solvent extraction method. *J. Microencapsul.* 2000, 17 (5), 587-598.
- Kas, H.S. Chitosan: properties, preparations and application to microparticulate systems. *J. Microencapsul.* 1997, 14 (6), 689-711.
- Kumar, S.G.; Kulkarni, A.R.; Aminabhavi, T.M. Cross-linked chitosan microspheres for encapsulation of diclofenac sodium: effect of crosslinking agent. *J. Microencapsul.* 2002, 19 (2), 173-180.
- Kim, S.E.; Park, J.H.; Cho, Y.W.; Chung, H.; Jeong, S.Y.; Lee, E.B.; Kwon, J.C. Porous chitosan scaffold containing microspheres loaded with transforming growth factor-beta: implications for cartilage tissue engineering. *J. Controlled Release* 2003, 91, 365-374.
- Yun, Y.H.; Goette, D.J.; Yellen, P.; Chen, W. Hyaluronic microspheres for sustained gene delivery and site-specific targeting. *Biomaterials* 2004, 25 (1), 147-157.
- Chickering, D.E.; Jacob, J.S.; Desai, T.A.; Harrison, M.; Harris, V.P.; Morrell, C.N.; Chaturvedi, P.; Mathiowetz, E. Biodegradable microspheres: III. An in vivo transit and bioavailability study of drug-loaded alginate and poly(lactide-co-glycolide anhydride) microspheres. *J. Controlled Release* 1997, 46, 35-46.
- Whateley, T.L. Microcapsules: preparation by interfacial polymerization and interfacial complexation and their application. In *Microencapsulation: Methods and Industrial Applications*; Benita, S., Ed.; Dekker: New York, U.S.A., 1996; Vol. 73, 349-375.
- Ting, T.; Gonda, I.; Gripps, E.M. Microparticles of PVA for nasal delivery. I. Generation by spray-drying and spray-desolvation. *Pharm. Res.* 1992, 9 (10), 1330-1335.
- Sam, A.P.; Hsiao, F.D.; Dixit, C. A Novel Process for Manufacturing PLG Microparticles by Spray Desolvation Avoiding the use of Toxic Solvents. International Symposium on Controlled Release of Bioactive Materials. Nice, France, 1994; 198-199.

31. Gombotz, W.R.; Healy, M.S.; Brown, L.R. Very Low Temperature Casting of Controlled Release Microspheres. U.S. Patent 5019400, 1991.
32. Johnson, O.L.; Cleland, J.L.; Lee, H.J.; Charnis, M.; Duenas, E.; Jaworowicz, W.; Shepard, D.; Shahzamani, A.; Jones, A.J.; Putney, S.D. A month-long effect from a single injection of microencapsulated human growth hormone. *Nat. Med.* 1996, 2, 795-799.
33. Lee, H.J.; Riley, G.; Johnson, O.; Cleland, J.L.; Kim, N.; Charnis, M.; Bailey, L.; Duenas, E.; Shahzamani, A.; Marian, M.; Jones, A.J.; Putney, S.D. In vivo characterization of sustained-release formulations of human growth hormone. *J. Pharmacol. Exp. Ther.* 1997, 281 (3), 1431-1439.
34. Santos, L.R.D.; Richard, J.; Pech, B.; Thies, C.; Benoit, J.P. Microencapsulation of protein particles within lipids using a novel supercritical fluid process. *Int. J. Pharm.* 2002, 242 (1-2), 69-78.
35. Young, T.J.; Johnston, K.P.; Mishima, K.; Tanaka, H. Encapsulation of lysozyme in a biodegradable polymer by precipitation with a vapor-over-liquid antisolvent. *J. Pharm. Sci.* 1999, 88 (6), 640-650.
36. Sunkara, G.; Kompella, U.B. Drug delivery applications of supercritical fluid technology. *Drug Delivery Technol.* 2002, 44 (1), 46-50.
37. Ogawa, Y.; Yamamoto, M.; Okada, H.; Yashiki, T.; Shimamoto, T. A new technique to efficiently entrap leuprolide acetate into microcapsules of poly(lactic acid) or copoly(lactic/glycolic) acid. *Chem. Pharm. Bull.* 1988, 36 (3), 1095-1103.
38. Viswanathan, N.B.; Thomas, P.A.; Pandit, J.K.; Kulkarni, M.G.; Mashelkar, R.A. Preparation of non-porous microspheres with high entrapment efficiency of proteins by a (water-in-oil)-in-oil emulsion technique. *J. Controlled Release* 1999, 58 (1), 3-20.
39. Lim, S.T.; Martin, G.P.; Berry, D.J.; Brown, M.B. Preparation and evaluation of the in vitro drug release properties and mucoadhesion of novel microspheres of hyaluronic acid and chitosan. *J. Controlled Release* 2000, 66, 281-292.
40. Takka, S.; Acaruturk, F. Calcium alginate microparticles for oral administration. I. Effect of sodium alginate type on drug release and drug entrapment efficiency. *J. Microencapsul.* 1999, 16 (3), 275-290.
41. Takada, S.; Uda, Y.; Toguchi, H.; Ogawa, Y. Application of a spray drying technique in the production of TRH-containing injectable sustained-release microparticles of biodegradable polymers. *PDA J. Pharm. Sci. Technol.* 1995, 49 (4), 180-184.
42. Ruon, G.; Feng, S.-S. Preparation and characterization of poly(lactic acid)-poly(ethylene glycol)-poly(lactic acid) (PLA-PEG-PLA) microspheres for controlled release of paclitaxel. *Biomaterials* 2003, 24, 5037-5044.
43. Hickey, A.J.; Tian, Y.; Parnasmpur, D.; Kanke, M. Biliary elimination of bronsulfamide, phenolphthalein, and doxorubicin released from microspheres following intravenous administration. *Biopharm. Drug Disposition* 1993, 14 (2), 181-186.
44. Perez, C.; Castellanos, J.J.; Costantino, H.R.; Al-Azzam, W.; Griebowicz, K. Recent trends in stabilizing protein structure upon encapsulation and release from biodegradable polymers. *J. Pharm. Pharmacol.* 2002, 54 (3), 301-313.
45. Knezevic, Z.; Gosaki, D.; Hrstic, M.; Jalscjak, I. Fluid-bed microencapsulation of ascorbic acid. *J. Microencapsul.* 1998, 15 (2), 237-252.
46. Palmieri, G.P.; Lauri, D.; Martelli, S.; Wehrle, P. Methoxybutyrate microencapsulation by gelatin-acacia complex concentration. *Drug Dev. Ind. Pharm.* 1999, 25 (4), 399-407.
47. Okada, H.; Heya, T.; Ogawa, Y.; Shimamoto, T. One-month release injectable microcapsules of luteinizing hormone-releasing hormone agonist (leuprolide acetate) for treating experimental endometriosis in rats. *J. Pharmacol. Exp. Ther.* 1988, 244 (2), 744-750.
48. Zhu, G.; Mallory, S.R.; Schwendeman, S.P. Stabilization of proteins encapsulated in injectable PLGA. *Nat. Biotechnol.* 2000, 18, 52-57.
49. Walter, E.; Moelling, K.; Pavlovic, J.; Merkle, H.P. Microencapsulation of DNA using poly(DL-lactide-co-glycolide): stability issues and release characteristics. *J. Controlled Release* 1999, 61 (3), 361-374.
50. Wang, C.; Ge, Q.; Ting, D.; Nguyen, D.; Shen, H.-r.; Chen, J.; Eisen, H.N.; Heller, J.; Langer, R.; Putnam, D. Molecularly engineered poly(ortho ester) microspheres for enhanced delivery of DNA vaccines. *Nat. Mater.* 2004, 3, 190-196.
51. Little, S.R.; Lynn, D.M.; Ge, Q.; Anderson, D.G.; Puram, S.V.; Chen, J.; Eisen, H.N.; Langer, R. Poly-beta amino ester-containing microcapsules enhance the activity of non-viral genetic vaccines. *Proc. Natl. Acad. Sci. U.S.A.* 2004, 101 (26), 9534-9539.
52. Schwendeman, S.P. Recent advances in the stabilization of proteins encapsulated in injectable PLGA delivery systems. *Crit. Rev. Ther. Drug Carrier Syst.* 2002, 19 (1), 73-98.
53. van de Weert, M.; Hennink, W.E.; Jiskoot, W. Protein instability in PLGA microparticles. *Pharm. Res.* 2000, 17 (10), 1159-1167.
54. Sah, H. Protein instability toward organic solvent/water emulsification: implications for protein microencapsulation into microspheres. *PDA J. Pharm. Sci. Technol.* 1999, 53 (1), 3-10.
55. Lu, W.; Park, T.G. Protein release from poly(lactic-co-glycolic acid) microspheres: protein stability problems. *PDA J. Pharm. Sci. Technol.* 1995, 49 (1), 13-19.
56. Fu, K.; Pack, D.W.; Kilbanov, A.M.; Langer, R. Visual evidence of acidic environment within degrading poly(lactic-co-glycolic acid) (PLGA) microspheres. *Pharm. Res.* 2000, 17 (1), 100-105.
57. Manning, M.C.; Patel, K.; Borchardt, R.T. Stability of protein pharmaceuticals. *Pharm. Res.* 1989, 6 (11), 903-918.
58. Shao, P.G.; Bailey, L.C. Porcine insulin biodegradable polyester microspheres: stability and in vitro release characteristics. *Pharm. Dev. Technol.* 2000, 3 (1), 1-9.
59. Lucke, A.; Kiermair, J.; Gopfrich, A. Peptide acylation by poly(alphahydroxy esters). *Pharm. Res.* 2002, 19 (2), 175-181.
60. Crotts, G.; Park, T.G. Stability and release of bovine serum albumin encapsulated within PLGA microparticles. *J. Controlled Release* 1997, 44, 123-134.
61. Shukry, V.; Kilbanov, A.M.; Langer, R. Mechanism of insulin aggregation and stabilization in agitated aqueous solutions. *Biotechnol. Bioeng.* 1992, 40, 895-903.
62. Yoo, Y.; Park, K. Microencapsulation of protein drugs: a novel approach. In *Biomaterials Handbook—Advanced Applications of Basic Sciences and Bioengineering*; Wise, D.L., Hsai, V., Lewandowski, K.-U., Yaszemski, M.J., Atabelli, D.E., Trantolo, D.J., Eds.; Marcel Dekker, Inc.: 2004; 305-332.
63. Sah, H. Stabilization of proteins against methylene chloride/water interface-induced denaturation and aggregation. *J. Controlled Release* 1999, 58 (2), 143-151.
64. Sah, H.; Smith, M.; Chern, R. A novel method of preparing PLGA microcapsules utilizing methyl ethyl ketone. *Pharm. Res.* 1996, 13 (3), 360-367.
65. Jiang, W.; Schwendeman, S.P. Stabilization and controlled release of bovine serum albumin encapsulated in poly(DL-lactide) and poly(ethylene glycol) microsphere blends. *Pharm. Res.* 2001, 18 (6), 878-885.
66. Yang, Y.Y.; Wan, J.P.; Chung, T.S.; Pallathadka, P.K.; Ng, S.; Heller, J. POE-PEG-POE triblock copolymeric microspheres containing protein I. Preparation and characterization. *J. Controlled Release* 2001, 75 (1-2), 115-128.
67. Bouillot, P.; Ubrich, N.; Sommer, F.; Dne, T.M.; Loeffler, J.P.; Dellachrie, E. Protein encapsulation in biodegradable amphiphilic microspheres. *Int. J. Pharm.* 1999, 181 (2), 159-172.

68. Wang, N.; Wu, X.S.; Li, J.K. A heterogeneously structured composite based on poly(lactide-co-glycolic acid) microspheres and poly(vinyl alcohol) hydrogel nanoparticles for long-term protein drug delivery. *Pharm. Res.* 1999, 16 (9), 1439-1455.
69. Blanco, M.D.; Alonso, M.J. Development and characterization of protein-loaded poly(lactide-co-glycolide) microspheres. *Eur. J. Pharm. Biopharm.* 1997, 43, 287-294.
70. Yeo, Y.; Chen, A.U.; Basaran, O.A.; Park, K. Solvent exchange method: a novel microencapsulation technique using dual microdispensers. *Pharm. Res.* 2004, 21 (8), 1419-1427.
71. Yeo, Y.; Basaran, O.A.; Park, K. A new process for making reservoir-type microcapsules using ink-jet technology and interfacial phase separation. *J. Controlled Release* 2003, 93 (2), 161-173.
72. Kumar, N.; Langer, R.S.; Domb, A.J. Polyhydrides: an overview. *Adv. Drug Delivery Rev.* 2002, 54 (7), 889-910.
73. Heller, J.; Barr, J.; Ng, S.Y.; Abdellau, K.S.; Gurny, R. Poly(ortho ester)s: synthesis, characterization, properties and uses. *Adv. Drug Delivery Rev.* 2002, 54 (7), 1015-1039.
74. Heller, J. Poly(ortho ester)s—some recent developments. *Polym. Mater. Sci. Eng.* 2003, 89, 189.
75. Bai, X.-L.; Yang, Y.-Y.; Chung, T.-S.; Ng, S.; Heller, J. Effect of polymer compositions on the fabrication of poly(ortho-ester) microspheres for controlled release of protein. *J. Appl. Polym. Sci.* 2001, 80, 1630-1642.
76. Slivnik, R.; Domb, A.J. Stereocomplexes of manitolomic lactic acid and sebacic acid ester-anhydride triblock copolymers. *Biomacromolecules* 2002, 3, 754-760.
77. Igartua, M.; Hernandez, R.M.; Esquisabel, A.; Gascon, A.R.; Calvo, M.B.; Pedraz, J.L. Influence of formulation variables on the in-vivo release of albumin from biodegradable microparticulate systems. *J. Microencapsul.* 1997, 14 (3), 349-356.
78. Huang, X.; Brazel, C.S. On the importance and mechanisms of burst release in matrix-controlled drug delivery systems. *J. Controlled Release* 2001, 73, 121-136.
79. Yang, Y.-Y.; Chung, T.-S.; Ping Ng, N. Morphology, drug distribution, and in vitro release profiles of biodegradable polymeric microspheres containing protein fabricated by double-emulsion solvent extraction/evaporation method. *Biomaterials* 2001, 22 (3), 231-241.
80. Pekarek, K.J.; Jacob, J.S.; Mathiowitz, E. Double-walled polymer microspheres for controlled drug release. *Nature* 1994, 367, 258-260.
81. Mathiowitz, E.; Langer, R. Multivall Polymeric Microspheres. U.S. Patent 5912017, 1999.
82. Berklind, C.; Pollauf, E.; Pack, D.W.; Kim, K.K. Uniform double-walled polymer microspheres of controllable shell thickness. *J. Controlled Release* 2004, 96, 101-111.
83. Gopferich, A.; Alonso, M.J.; Langer, R. Development and characterization of microencapsulated microspheres. *Pharm. Res.* 1994, 11 (11), 1568-1574.
84. Wang, H.T.; Schmitt, E.; Flanagan, D.R.; Linhardt, R.J. Influence of formulation methods on the in vitro controlled release of protein from poly(ester) microspheres. *J. Controlled Release* 1991, 17, 23-32.
85. Yamaguchi, Y.; Takemura, M.; Kitagawa, A.; Ogawa, Y.; Mizushima, Y.; Iguchi, R. Insulin-loaded biodegradable PLGA microspheres: initial burst release controlled by hydrophilic additives. *J. Controlled Release* 2002, 81 (3), 235-249.
86. Li, X.; Deng, X.; Huang, Z. In vitro protein release and degradation of poly(D,L-lactide-poly(ethylene glycol) microspheres with entrapped human serum albumin. *Pharm. Res.* 2001, 18 (1), 117-124.
87. Sah, H.K.; Toddysala, R.; Chien, Y.W. Biodegradable microcapsules prepared by a w/o/w technique: effects of shear force to make a primary w/o emulsion on their morphology and protein release. *J. Microencapsul.* 1995, 12 (1), 59-69.
88. Yan, C.; Resau, J.H.; Hewison, J.; Mest, M.; Rill, W.L.; Kende, M. Characterization and morphological analysis of protein-loaded poly(lactide-co-glycolide) microspheres prepared by water-in-oil-in-water emulsion technique. *J. Controlled Release* 1994, 32, 231-241.
89. Yang, Y.-Y.; Chia, T.-H.; Chung, T.-S. Effect of preparation temperature on the characteristics and release profiles of PLGA microspheres containing protein fabricated by double-emulsion solvent extraction/evaporation method. *J. Controlled Release* 2000, 69 (1), 81-96.
90. Sah, H.; Toddysala, R.; Chien, Y.W. The influence of biodegradable microcapsule formulations on the controlled release of a protein. *J. Controlled Release* 1994, 30, 201-211.
91. Walter, E.; Dreher, D.; Kok, M.; Thiele, L.; Kiama, S.G.; Gehr, P.; Merkle, H.P. Hydrophilic poly(D,L-lactide-co-glycolide) microspheres for the delivery of DNA to human-derived macrophages and dendritic cells. *J. Controlled Release* 2001, 76 (1-2), 149-168.
92. Hsu, Y.-Y.; Hao, T.; Hedley, M.L. Comparison of process parameters for microencapsulation of plasmid DNA in poly(D,L-lactide-co-glycolide) acid microspheres. *J. Drug Targeting* 1999, 7 (4), 313-323.
93. Jiang, G.; Thanoo, B.C.; DeLuca, P.P. Effect of osmotic pressure in the solvent extraction phase on BSA release profile from PLGA microspheres. *Pharm. Dev. Technol.* 2002, 7 (4), 391-399.
94. Narayani, R.; Rao, K.P. Gelatin microsphere cocktails of different sizes for the controlled release of anticancer drugs. *Int. J. Pharm.* 1996, 143, 255-258.
95. Bezeiner, J.M.; Radersma, R.; Grijpma, D.W.; Dijkstra, P.J.; Blitterswijk, C.A.; Feijen, J. Microspheres for protein delivery prepared from amphiphilic multiblock copolymers. 2. Modulation of release rate. *J. Controlled Release* 2000, 67, 249-260.
96. Berklind, C.; King, M.; Cox, A.; Kim, K.; Pack, D.W. Precise control of PLG microsphere size provides enhanced control of drug release rate. *J. Controlled Release* 2002, 82, 137-147.
97. Berklind, C.; Kim, K.K.; Pack, D.W. Protein Release from Uniform Poly(D,L-lactide-co-glycolide) Microspheres. 31st International Symposium on Controlled Release of Bioactive Materials. Honolulu, Hawaii, 2004; pp. 472.
98. Amsden, B.G.; Goosen, M.F.A. An examination of factors affecting the size, distribution and release characteristics of polymer microbeads made using electrostatics. *J. Controlled Release* 1997, 43 (2-3), 183-196.
99. Amsden, B. The production of uniformly sized polymer microspheres. *Pharm. Res.* 1999, 16 (7), 1140-1143.
100. Berklind, C.; Kim, K.; Pack, D.W. Fabrication of PLG microspheres with precisely controlled and monodisperse size distributions. *J. Controlled Release* 2001, 73, 59-74.
101. Mathiowitz, E.; Jacob, J.S.; Jang, Y.S.; Carino, G.P.; Chickering, D.E.; Chaturvedi, P.; Santos, C.A.; Vijayaraghavan, K.; Montgomery, S.; Bassett, M.; Morrell, C. Biologically erodable microspheres as potential oral drug delivery systems. *Nature* 1997, 386, 410-414.
102. Santos, C.A.; Freedman, B.D.; Ghosh, S.; Jacob, J.S.; Scarpulla, M.; Mathiowitz, E. Evaluation of anhydride oligomers within polymer microsphere blends and their impact on bioadhesion and drug delivery in vitro. *Biomaterials* 2003, 24, 3571-3583.
103. Thanos, C.G.; Liu, Z.; Goddard, M.; Reimeke, J.; Bailey, N.; Cross, M.; Buerli, R.; Mathiowitz, E. Enhancing the oral bioavailability of the poorly soluble drug diclofenac with a bioadhesive polymer. *J. Pharm. Sci.* 2003, 92 (8), 1677-1689.
104. Lynn, D.M.; Langer, R. Degradable poly(beta-amino ester)s: synthesis, characterization, and self-assembly with plasmid DNA. *J. Am. Chem. Soc.* 2000, 122, 10,761-10,768.